

Statistical Analysis of Molecular Data Using Software Packages



Lecture Notes on
Statistical analysis of genetic association studies among
unrelated individuals

First edition

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ABOUT

These lecture notes on “*Statistical analysis of genetic association studies among unrelated individuals*” were prepared under my lecture series on “*Statistical Analysis of Molecular Data Using Software Packages*”. This lecture was delivered to trainees attending 14 day training on “Application of Molecular and Bioinformatics Tools in Agriculture and Allied Sciences” organized by CST, UP Centre of Excellence in Agriculture Biotechnology in Collaboration with DBT funded Bioinformatics Infrastructure Facility, College of Biotechnology, SVPUAT during 7-20th July 2021. I have tried to explain how molecular data of a population can be analyzed using SPSS and HW_Test software packages. Overall, population association studies in which unrelated individuals for different economic traits are typed at a number of Single Nucleotide Polymorphism (SNP) markers excluding family-based association studies, admixture mapping or linkage studies have been introduced and explained. Use of diagrams, explanatory boxes, examples and tables have deliberately been used throughout the notes to create an interest among the trainees. Once through with these lecture notes readers will be able to understand the basics and application of statistical analysis of genetic association studies among unrelated individuals using statistical packages. I had tried my level best to simplify the concept in easy to understand language. Further constructive suggestions to improve this lecture notes are always welcome from readers on my email and whatsapp.

[KULDEEP KUMAR TYAGI]

DISCLAIMER

These lecture notes on “*Statistical analysis of genetic association studies among unrelated individuals*” under the lecture series on “*Statistical Analysis of Molecular Data Using Software Packages*” have been compiled from various resources available in the public domain. Only excerpts from the original works have been used. This is being done for educational purposes in the interest of developing a concise and updated reading material for students and trainees with no intent of commercial benefits. References to the source of material used have been included in the footnotes. The author does not claim any ownership of any copyrighted material included by chance in the lecture. If due to inability to trace the original source any copyrighted material got included, it may please be brought to the notice of the author for rectification.

[KULDEEP KUMAR TYAGI]

OTHER LECTURE NOTES BY THE AUTHOR

As on: 13-07-2021

Series: Animal Genetics & Breeding							
S.No.	Title	Date of Publication	Editions	Pages	Views	Down loads	Access
1	Introduction and importance of statistics and biostatistics	12-10-2020	First	20	40.1k	254	Download
2	Probability	20-08-2020	First	33	12.6k	193	Download
3	Probability	24-11-2020	Second	24	29.5k	280	Download
4	Probability Distribution	07-12-2020	First	33	12.6k	193	Download
5	Introduction of population genetics	23-02-2021	First	16	22.8k	128	Download
6	Genetic structure of population	23-02-2021	First	40	22.2k	211	Download
7	Hardy Weinberg Law	10-07-2021	First	42	49	0	Download

ACKNOWLEDGEMENT

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[KULDEEP KUMAR TYAGI]

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Statistical analysis of genetic association studies among unrelated individuals

People all around the world have been conducting genetic association studies for a long time. Still the literature to practically understand the statistical analysis of such data using software packages is scarce. Statistical analysis of molecular data is a vast topic. So we will be undertaking one of the important basic and preliminary topics “*Statistical analysis of genetic association studies among unrelated individuals*” in this lecture. You will be able to analyse molecular data of a population using SPSS and HW_Test software packages after going through this lecture. Overall, we will be discussing population association studies in which unrelated individuals for different economic traits are typed at a number of Single Nucleotide Polymorphism (SNP) markers excluding family-based association studies, admixture mapping or linkage studies.

1. Coding the genotypes

To demonstrate practical hands on we will be making use of dummy data. This dummy data is available in Excel workbook and can be [downloaded online](#). This data sheet represents SNP data of six genes say (Gene 1, Gene 2, Gene 3, Gene 4, Gene 5 and Gene 6) on 550 lactating buffaloes and their associated data on traits of economic importance like Age at First Calving (AFC) in days, Service Period (SP) in days and Standard 300 day Lactation yield (SLY) in Kg.

1.1. Feeding data in excel workbook

First data can be feeded in Excel workbook for the ease of data feeding. The header row will define the variables. The first column can be used to either assign the serial number or identification number of the buffalo. Columns thereafter constitute the variables like various genes and traits of economic importance. Data pertaining to each individual is assigned in a single row. Say for example serial number 101, the genotypes (coding) were AB(2), AA(1), AB(2), CD(9), AB(2), AB(2) for Gene 1, Gene 2, Gene 3, Gene 4, Gene 5 and Gene 6 respectively. Whereas the values for corresponding traits of economic importance Age at First Calving (AFC) is 1077 days, Service Period (SP) is 121 days and

Standard 300 day Lactation yield (SLY) is 2225 Kg respectively. Similarly, data pertaining to each buffalo is feeded into the excel workbook. Save your excel workbook on your computer in a folder designated for this analysis.

Table 1: Number of buffaloes pertaining to each genotype and their corresponding coding.

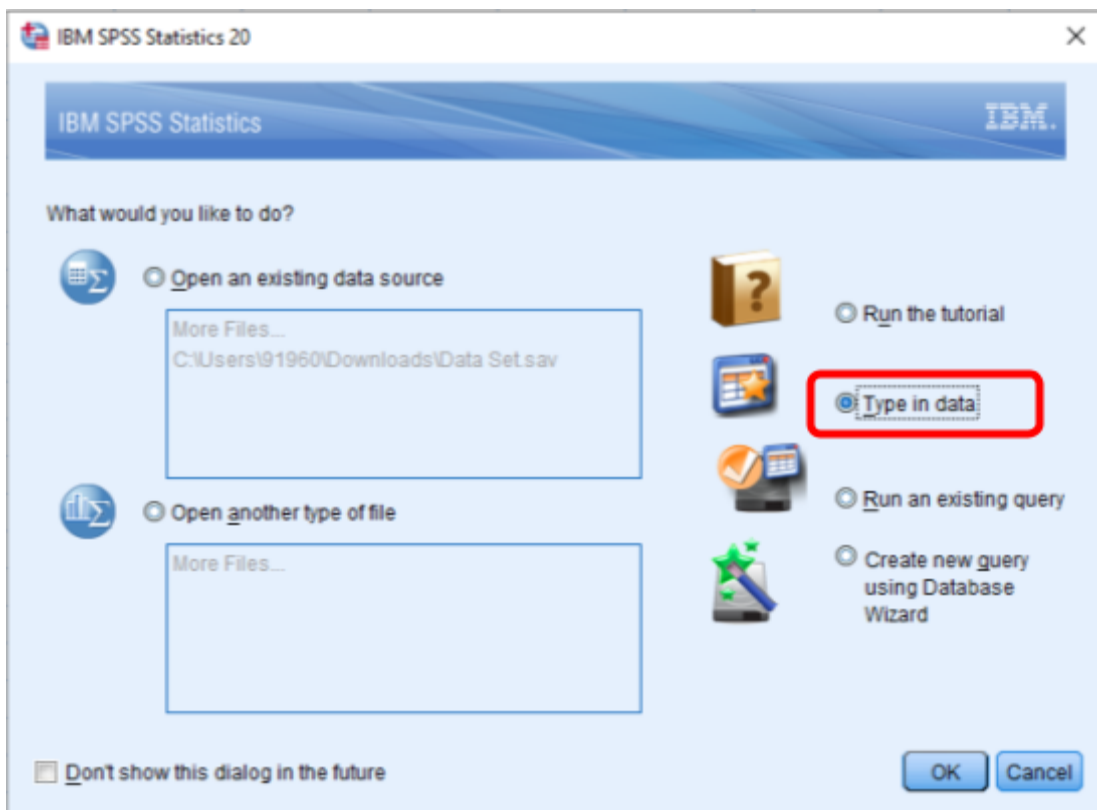
Genes		Number of buffaloes for each genotype				Coding for each genotype						
Gene 1	Two alleles		A	B		A	B					
	N	A	110	266		A	1	2				
	550	B				B		3				
Gene 2	One Allele		A			A						
	N	A	550			A	1					
	550											
Gene 3	Three Alleles		A	B	C		A	B	C			
	N	A	47	129	115	A	1	2	3			
	550	B		77	119	B		4	5			
		C				63	C			6		
Gene 4	Four Alleles		A	B	C	D		A	B	C	D	
	N	A	19	84	59	67	A	1	2	3	4	
	550	B		48	94	75	B		5	6	7	
		C				29	51	C			8	9
		D					24	D				10
Gene 5	Two Alleles		A	B			A	B				
	N	A	107	264			A	1	2			
	550	B				179	B		3			

Gene 6 Three Alleles		A	B	C				
N	A	36	174	177	A	1	2	3
550	B		28	101	B		4	5
	C			34	C			6

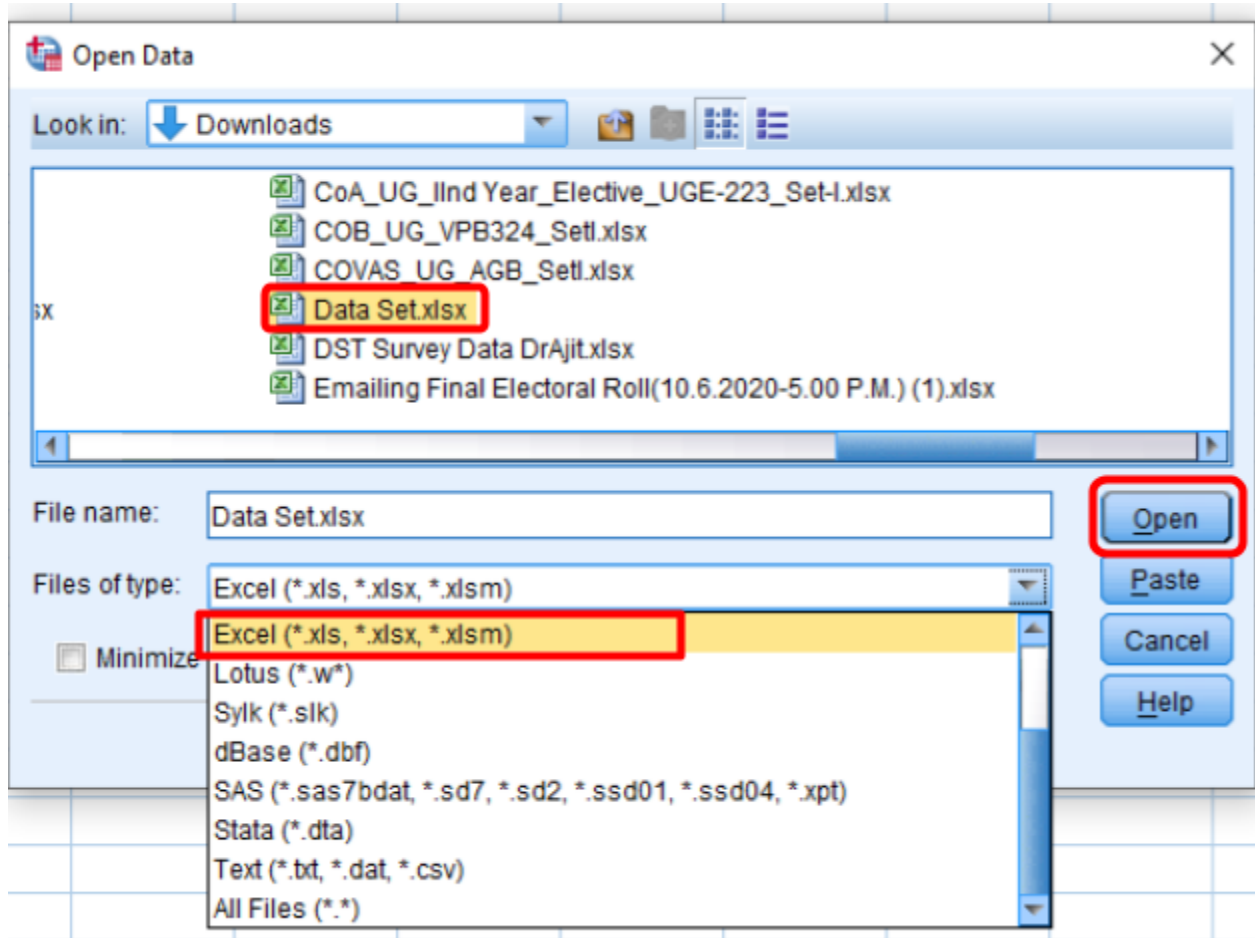
2.SPSS data sheet

2.2. Importing data to SPSS

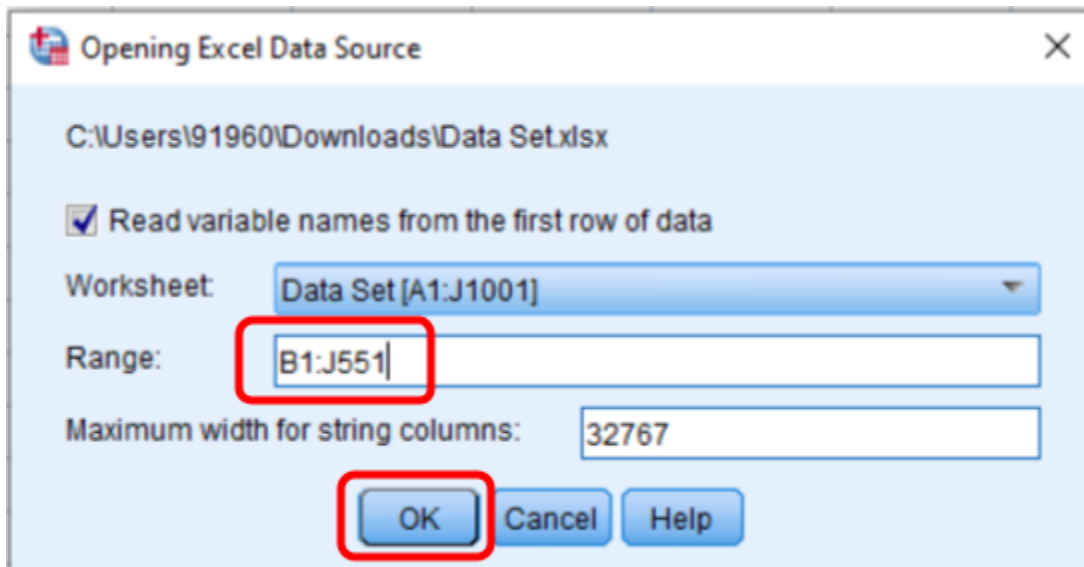
We will be importing data to SPSS once our excel workbook is ready. Open SPSS program installed on your computer. A dialog box as shown in the figure below will open.



Click on type in data to open the SPSS data view. Go to *File* ⇒ *Read Text Data*..It will prompt you to locate your data file as shown below.

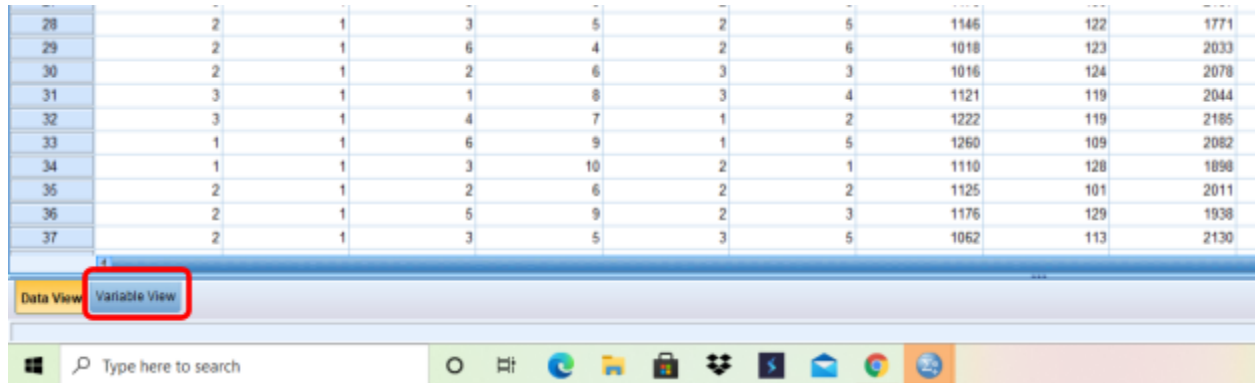


Select the File type from the drop down menu and then locate your file and click open. This will open another dialog box as shown below.

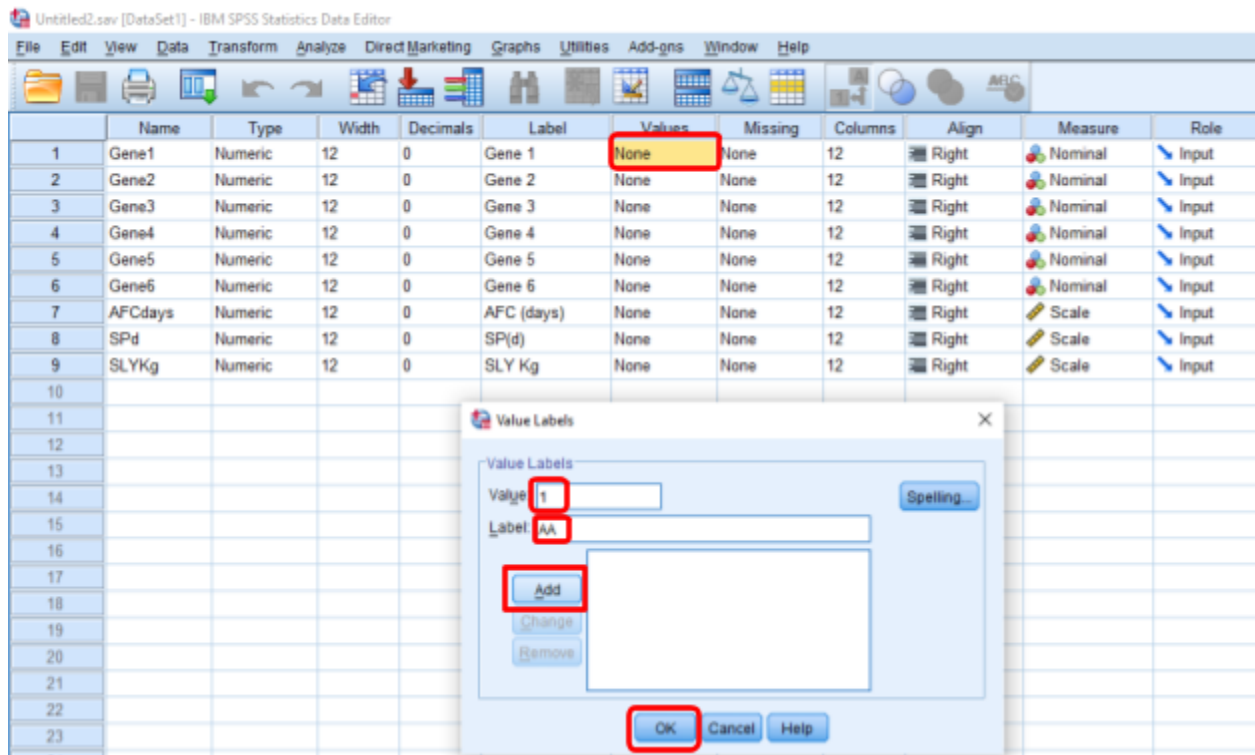


Define your data range (B1:J551) and then click OK. This will import all your excel data

into the SPSS data view. Now we will be defining variables by clicking the variable view tab given at the bottom left of SPSS.



This will open the variable view of the SPSS as shown below.



Click in the cell of the column labeled “Values”. This will open a dialog box in which value and their associated genotype (Label) can be added. Click on Ok once you complete feeding all the genotype labels corresponding to a gene. The final dataset file can be [downloaded here](#).

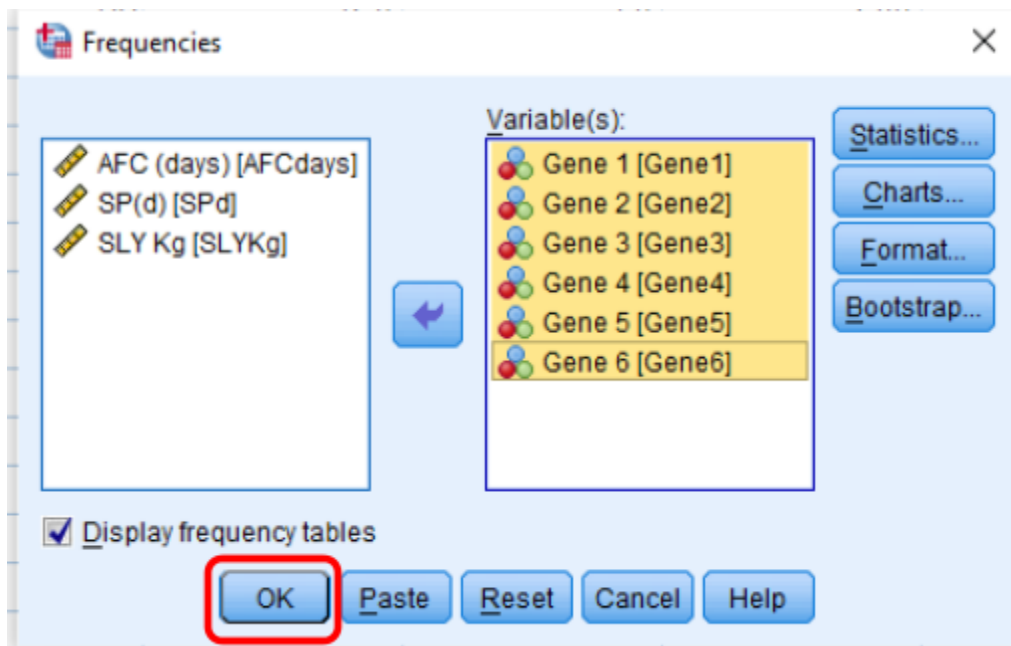
Now each genotype will be visible in the data view using the toggle button provided at the top menu ribbon as shown in the figure below. So basically if you consider gene 1, which has two alleles (A and B) will be having three genotypes AA, AB and BB and their

corresponding coding will be 1, 2 and 3 respectively. Similarly, coding patterns for other genes are also followed in a similar fashion. Coding our data in this way will help us to analyse our data with ease using above stated softwares.

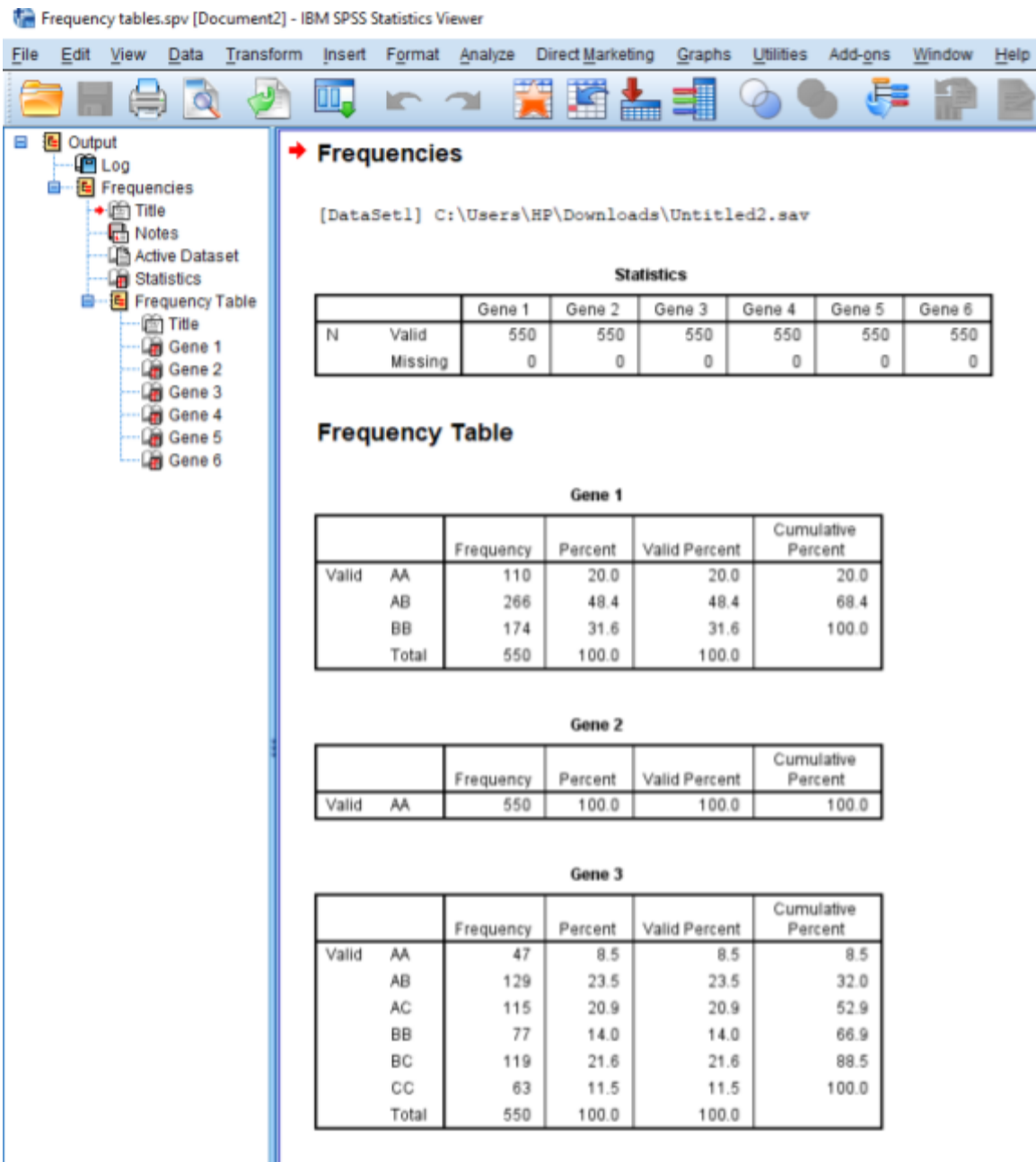
3. Analysis

3.1. Frequency table using SPSS

Go to *Analyze* ⇒ *Descriptive Statistics* ⇒ *Frequencies*. This will open a dialog box, select and put all the variable named genes into the variables box and click OK. This will give you an output file with frequency of genotypes for all the genes. The file can be [downloaded online](#)



Below is the screenshot of the output file. These frequencies are being feeded in table 1 and will be used subsequently in Hardy Weinberg equilibrium testing. The benefit of making this table is to have handy absolute frequencies for each genotype ready to be feeded in HW_TEST software. The format prevents mistakes while feeding the value of each genotype because it simulates the data feeding design available in the HW_TEST software.



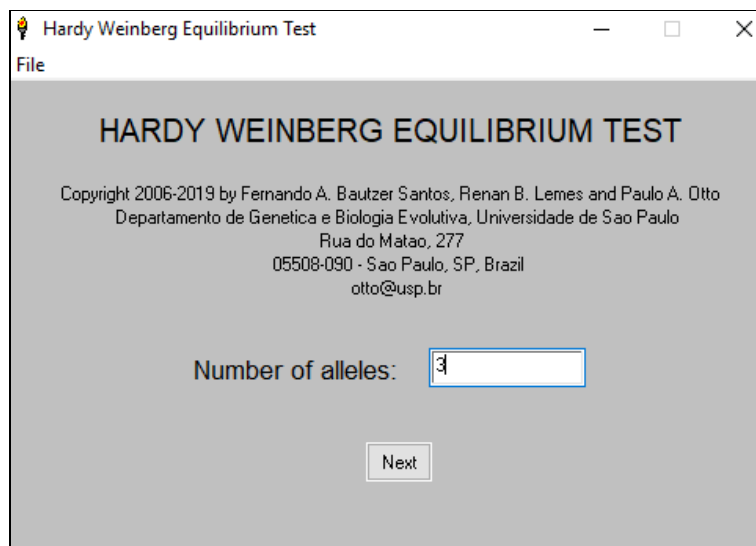
3.2. Testing for Hardy Weinberg Equilibrium (HWE)

Molecular data on a population are generally subjected to be tested for Hardy Weinberg equilibrium (HWE) as a measure to find preliminary anomalies. HWE test is used extensively on a routine basis to exclude samples with gross molecular typing defects from the usually very large sets of genetic markers presently used in various types of population genetic analyses. Natural or domesticated populations are generally

subjected to inbreeding, population stratification, selection or sometimes disease association may lead to deviations from HWE. Sometimes deviation from HWE may be an indication of presence of a common deletion polymorphism, a mutant PCR-primer site or because of a tendency to miscall heterozygotes as homozygotes. So far researchers¹ have tested for HWE primarily as a data quality check and have discarded loci that, for example, deviate from HWE among controls at significance level $\alpha = 10^{-3}$ or 10^{-4} . However, the possibility that a deviation from HWE is due to a deletion polymorphism or a segmental duplication that could be important in disease causation should now be considered before discarding loci.

We will be using user friendly, window based executable open source software HW_TEST for testing HWE². The program can be obtained free of charge directly from the github internet repository as an standalone executable zip file <https://github.com/Lemes-RenanB/HardyWeinbergTesting>. The downloadable zip file contains a well defined user manual. The Pearson test is easy to compute, but the χ^2 approximation can be poor when there are low genotype counts, and it is better to use a Fisher exact test, which does not rely on the χ^2 approximation.

3.2.1 Analysis using HW_TEST software



¹ Balding DJ. A tutorial on statistical methods for population association studies. Nat Rev Genet. 2006 Oct;7(10):781-91. doi: 10.1038/nrg1916. PMID: 16983374.

² Santos, F., Lemes, R. B., & Otto, P. A. (2020). HW_TEST, a program for comprehensive HARDY-WEINBERG equilibrium testing. Genetics and molecular biology, 43(2), e20190380. <https://doi.org/10.1590/1678-4685-GMB-2019-0380>

The opening window of software HW_TEST asks for the number of alleles in your data set. Let us consider a case of gene 3 which has three alleles. Put this value into the designated field and press next.

Now refer to the structural representation of gene 3 and their corresponding genotypes given in Table 1. The values corresponding to each genotype need to be filled in designated space provided in HW_TEST software.

The screenshot shows the Hardy Weinberg Equilibrium Test software interface. The window title is "Hardy Weinberg Equilibrium Test". The main title is "HARDY WEINBERG EQUILIBRIUM TEST". Under "Statistical Tests", the "Chi-square without correction" option is selected. Under "Number of Genotyped Individuals", the counts for alleles A, B, and C are entered as 47, 77, and 63 respectively. A "Run" button is located at the bottom of the interface.

	A	B	C
A	47		
B		77	
C			63

Once all the values are entered correctly, click on "Run". This will open the output file with results. The first result output provides us an idea about the observed and expected genotypes frequencies. One can see that the sum of these genotypic frequencies equals unity. Now, the second output provides us with the estimates of allelic frequencies. Finally the P value for χ^2 estimate is provided. If the value of "P" is greater than 0.05 indicates that our null hypothesis of no difference between observed and expected genotypic frequencies is accepted. Therefore, for the given gene the population is in

Hardy Weinberg equilibrium. We can assume for the given gene evolutionary forces like mutation, migration and selection are not operating in the given population at the time of screening.

Hardy-Weinberg Equilibrium Testing

File Edit

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 Departamento de Genética e Biologia Evolutiva
 Universidade de São Paulo
 Rua do Matao, 277
 05508-090 São Paulo SP, Brazil
 otto@usp.br

	genotype absolute frequencies		genotype relative frequencies	
	observed	expected	observed	expected
AA	47	51.929	0.085	0.094
AB	129	123.524	0.235	0.225
AC	115	110.618	0.209	0.201
BB	77	73.456	0.140	0.134
BC	119	131.564	0.216	0.239
CC	63	58.989	0.115	0.107

N 550

allele	allele freq. (pi)	s.e.(pi) H0:{d,h,r}	s.e.(pi) H1:{p^2,2pq,q^2}
A	0.307	0.014	0.014
B	0.365	0.015	0.015
C	0.327	0.014	0.014

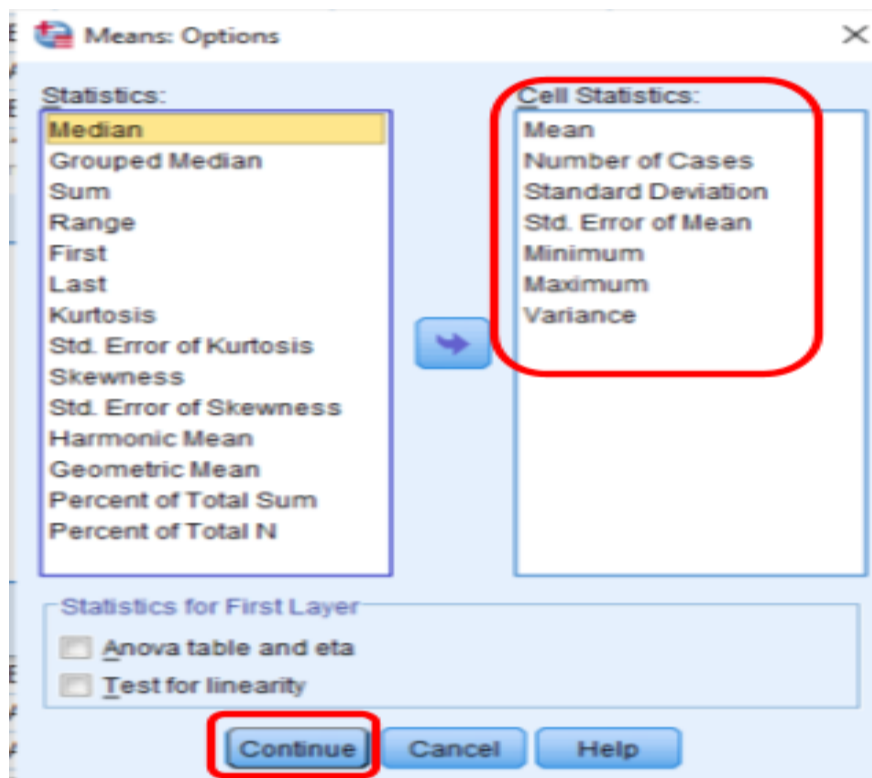
P(chi-squared test without correction) = 0.4683

3.3. Descriptive Statistics using SPSS

To obtain the descriptive statistics we click on Analyze followed by comparing means and then means ie. *Analyze* ⇒ *Comparing Means* ⇒ *Means*. This will open a dialog box. In the dialog box put all the traits of economic importance into the dependent variable box and all the genes into the independent list as shown in the figure given below.



Now this will open another dialog box in which various parameters for descriptive statistics can be defined.



Click on continue to save the changes and then Ok to obtain the results of descriptive statistics. The output file can be [downloaded here](#) for your ready reference to match your results when you practice this exercise.

Descriptive Statistics.spv [Document1] - IBM SPSS Statistics Viewer

File Edit View Data Transform Insert Format Analyze Direct Marketing Graphs Utilities

jt
 .log
 Means
 Title
 Notes
 Active Dataset
 Case Processing Summary
 AFC (days) SP(d) SLY Kg * Gen
 AFC (days) SP(d) SLY Kg * Gen
 AFC (days) SP(d) SLY Kg * Gen
 AFC (days) SP(d) SLY Kg * Gen
 AFC (days) SP(d) SLY Kg * Gen
 AFC (days) SP(d) SLY Kg * Gen

AFC (days) SP(d) SLY Kg * Gene 1

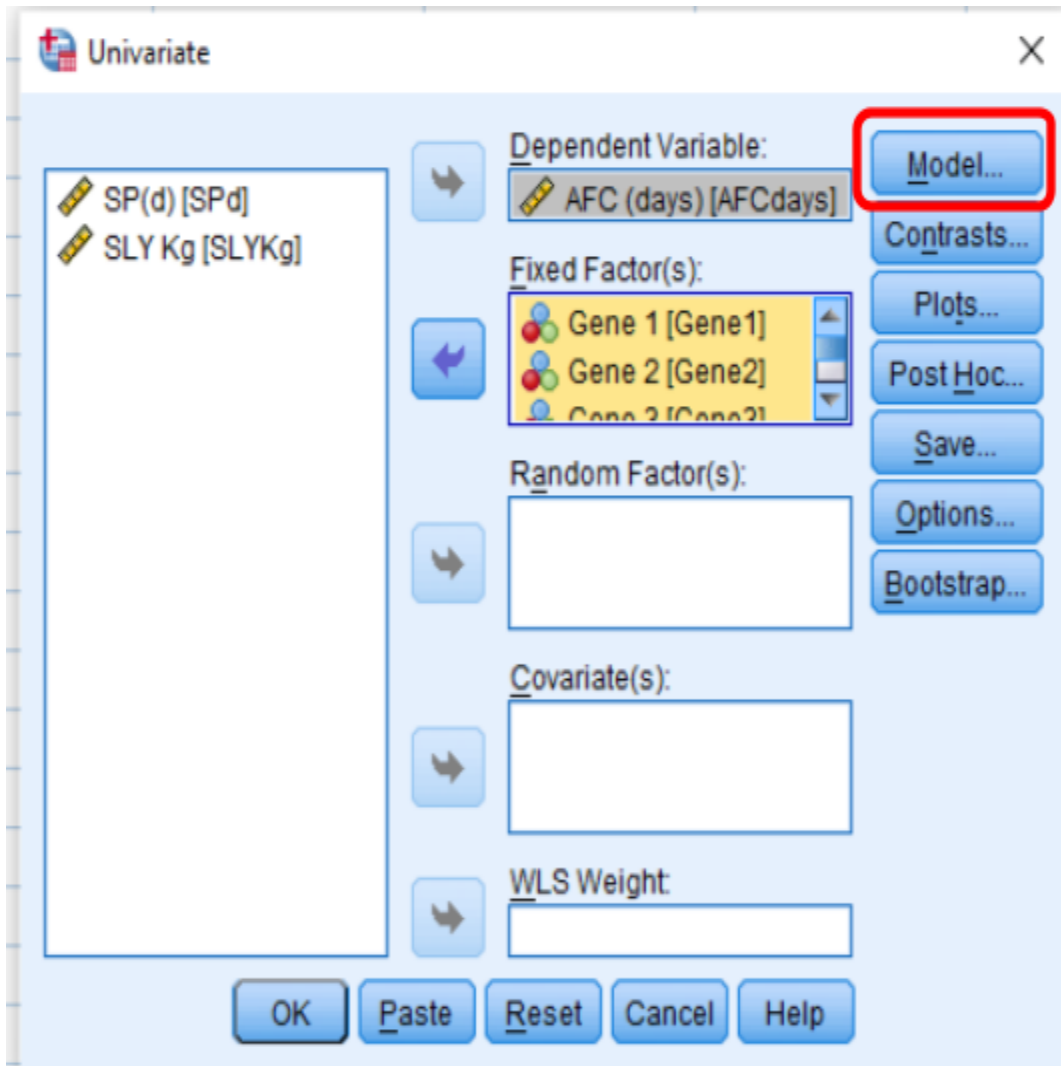
Gene 1		AFC (days)	SP(d)	SLY Kg
AA	Mean	1154.28	126.60	2065.25
	N	110	110	110
	Std. Deviation	98.570	10.494	140.987
	Std. Error of Mean	9.398	1.001	13.443
	Minimum	900	86	1688
	Maximum	1411	149	2518
	Variance	9716.039	110.114	19877.219
AB	Mean	1173.55	126.73	2072.97
	N	266	266	266
	Std. Deviation	97.540	11.303	153.680
	Std. Error of Mean	5.981	.693	9.423
	Minimum	949	86	1698
	Maximum	1423	158	2627
	Variance	9514.067	127.751	23617.535
BB	Mean	1194.81	126.26	2087.71
	N	174	174	174
	Std. Deviation	104.815	10.973	148.846
	Std. Error of Mean	7.946	.832	11.284
	Minimum	867	95	1680
	Maximum	1535	154	2537
	Variance	10986.224	120.412	22154.995
Total	Mean	1176.42	126.56	2076.09
	N	550	550	550
	Std. Deviation	100.959	11.023	149.663
	Std. Error of Mean	4.305	.470	6.382
	Minimum	867	86	1680
	Maximum	1535	158	2627

The output provides, mean, number of observations, standard deviation, standard error of mean, minimum, maximum and variance pertaining to each genotype.

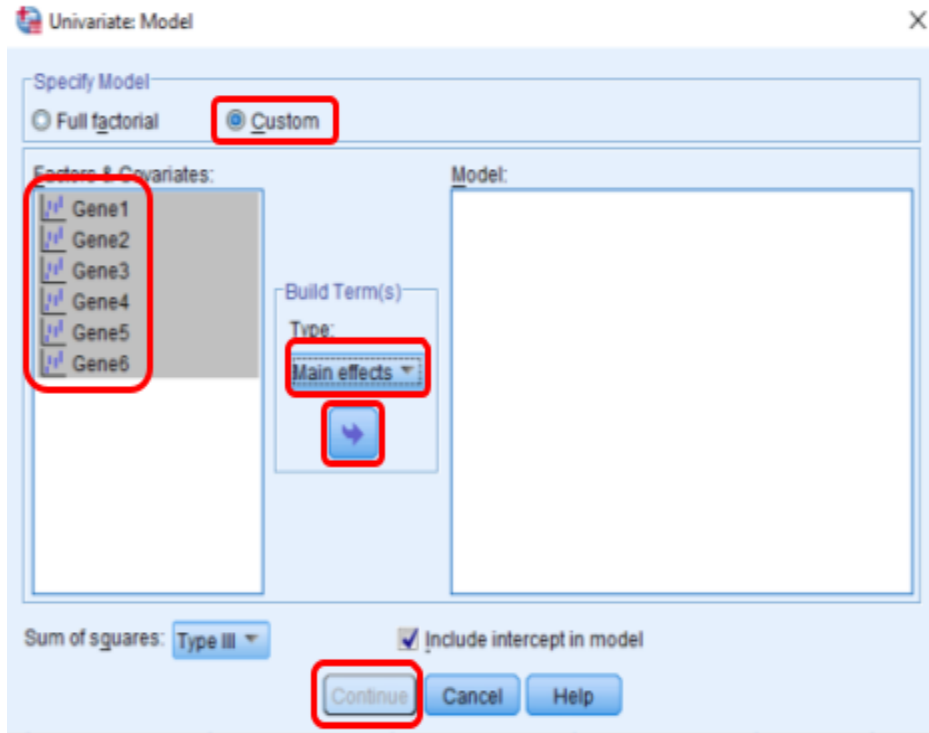
3.3. Analysis of Variance (ANOVA) and Duncan test using SPSS

Now we will be subjecting our data for finding any association between various genes studied and traits of economic importance available with us for each individual.

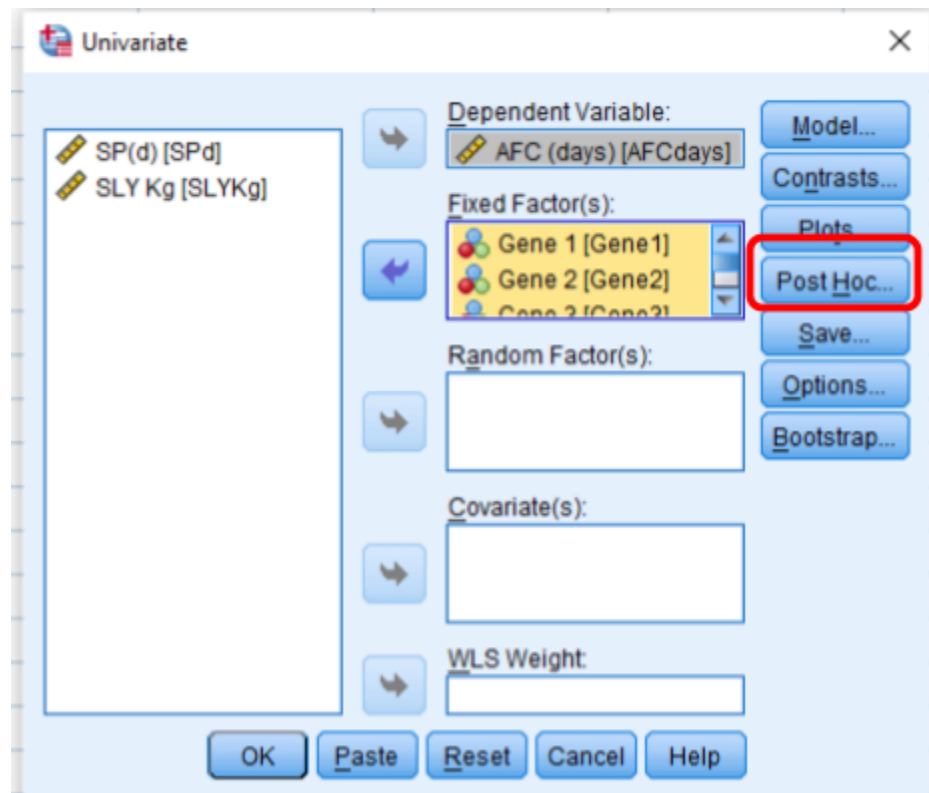
Go to *Analyze* ⇒ *General Linear Model* ⇒ *Univariate*. This will open another dialog box. Put one quantitative trait (say AFC) into the dependent variable box and all the genes in the Fixed factor box as shown below. Then click on the model which will open another dialog box to define our model for ANOVA.



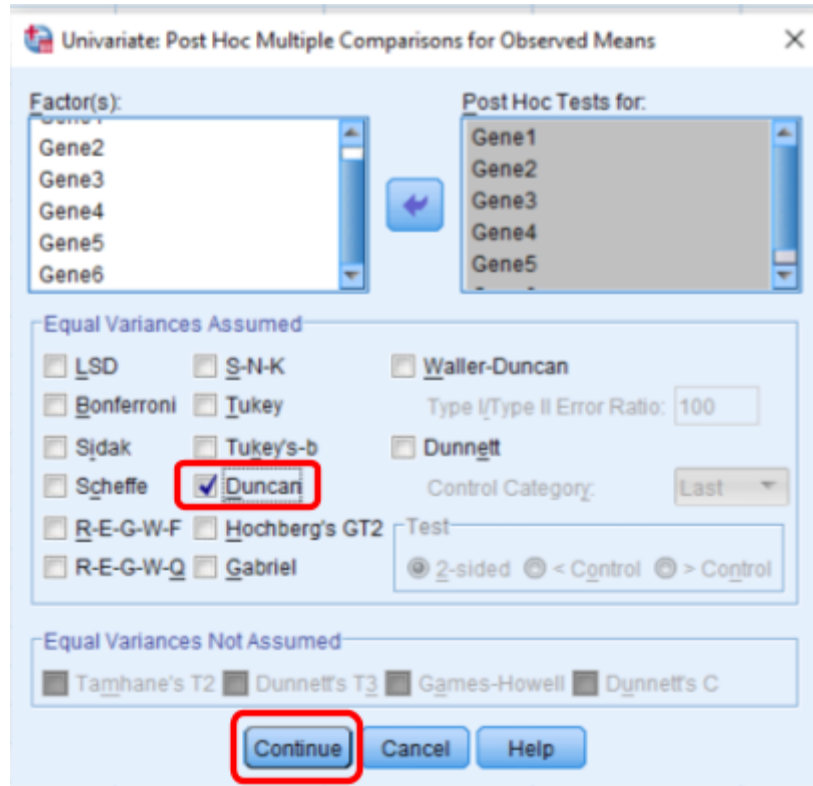
Click on the custom radio button, select all the fixed factors and choose main effects type from the drop down menu of build terms. Click on the arrow to put all fixed factors into the model box and to activate the continue button. Click on continue to save your changes.



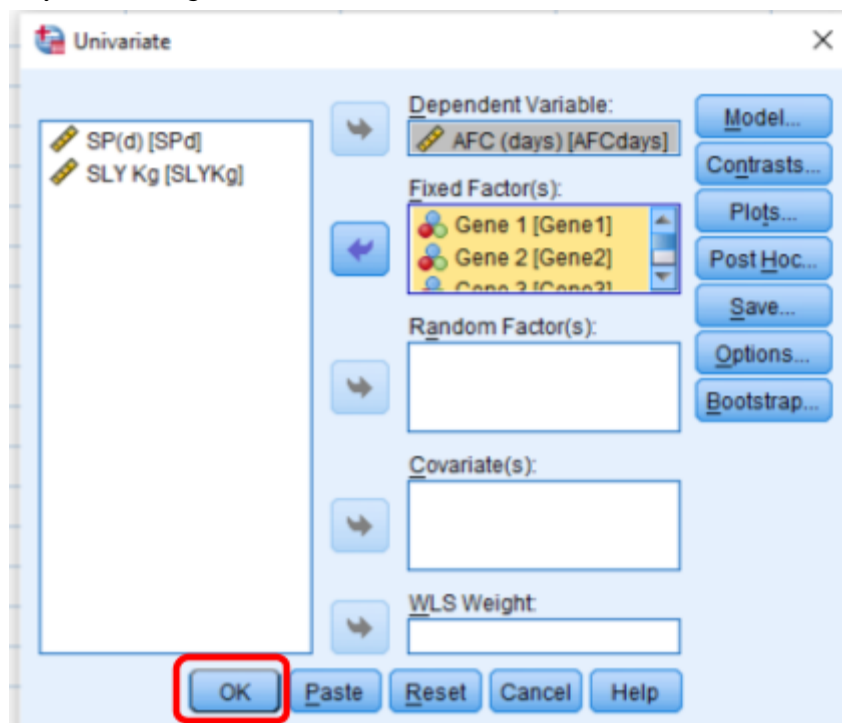
Now click on the Post Hoc button to carry out post hoc tests in your analysis.



This will open another dialog box as shown below.



Put all the fixed factors into the post hoc box, select Duncan checkbox and click on continue to save your changes. Now click on Ok to obtain the results.



This will open the output window with desired results. Similarly other two traits of economic importance will be put in the dependent variable box one by one and click ok to obtain the results. The output file can be [downloaded here](#) for your ready reference to match your results when you practice this exercise.

The first table we need to lookup in the output file is “Test of between subject effect”. This is the ANOVA table.

ANOVA.spv [Document2] - IBM SPSS Statistics Viewer

File Edit View Data Transform Insert Format Analyze Direct Marketing Graphs Utilities Add-ons Window Help

Output

- Log
- Univariate Analysis
 - Title
 - Notes
 - Active Dataset
 - Warnings
 - Between-Subjects Effects
 - Tests of Between-Subjects Effects
 - Post Hoc Tests
 - Gene 1
 - Title
 - Homo
 - T
 - A
 - Gene 3
 - Title
 - Homo
 - T
 - A
 - Gene 4
 - Title
 - Homo
 - T
 - A

Tests of Between-Subjects Effects

Dependent Variable: AFC (days)

Source	Type III Sum of Squares	df	Mean Square	F	Sig.
Corrected Model	235629.529 ^a	23	10244.762	1.005	.456
Intercept	304306685.0	1	304306685.0	29861.737	.000
Gene1	97386.253	2	48693.126	4.778	.009
Gene2	.000	0	.	.	.
Gene3	46937.857	5	9387.571	.921	.467
Gene4	27702.804	9	3078.089	.302	.974
Gene5	12412.451	2	6206.225	.609	.544
Gene6	30790.643	5	6158.129	.604	.697
Error	5360214.609	526	10190.522		
Total	766778406.0	550			
Corrected Total	5595844.138	549			

a. R Squared = .042 Adjusted R Squared = .000

The table shown above is for the dependent variable AFC. The first column shows the sources of variation which in our case are different genes. Now in this table we have to look for the source of variation which is significantly affecting our dependent variable. The gene 1 in this case is contributing a significant effect with $p = 0.009$ which is less than 0.05. No other gene is causing a significant effect on our dependent variable AFC. The R squared value of 0.042 represents that around 4.2 % variation in our dependent variable is explained by our model. Adjusted R-squared is a modified version of R-squared that has been adjusted for the number of predictors in the model. The adjusted R-squared increases when the new term improves the model more than would be expected by chance. It decreases when a predictor improves the model by less than expected.

Since only gene 1 is contributing significant effect on dependent variable the post-hoc tests pertaining to this gene will only be considered. The output results of post-hoc Duncan test are summarized under the heading of “Homogeneous Subsets”.

Gene 1

Homogeneous Subsets

AFC (days)

Duncan^{a,b,c}

Gene 1	N	Subset	
		1	2
AA	110	1154.28	
AB	266	1173.55	1173.55
BB	174		1194.81
Sig.		.087	.059

Means for groups in homogeneous subsets are displayed.
Based on observed means.
The error term is Mean Square(Error) = 10190.522.

- a. Uses Harmonic Mean Sample Size = 161.313.
- b. The group sizes are unequal. The harmonic mean of the group sizes is used. Type I error levels are not guaranteed.
- c. Alpha = .05.

The output table for Duncan test is quite helpful in determining the superscripts of means that differed significantly ($\alpha= 0.05$). Thus the mean AFC (days) for genotype AA, AB and BB will bear superscripts a, ab and b respectively. Similarly results for all the genes can be reported in similar fashion. This enables us to analyze molecular data for testing Hardy Weinberg equilibrium along the identification of genes affecting dependent variables significantly. The method also gives us an idea about the percent variation explained by our predictor variables in the model.

4. Summary

This lecture on “Statistical analysis of genetic association studies among unrelated individuals” taught us one of the methods of coding our genotype data. Thereafter data is feeded as each column representing a variable and each row representing observations made on an individual. It has been observed that HW_TEST software is user friendly and proved to be an indispensable tool for testing a population for Hardy Weinberg equilibrium. This software also provides us the estimates of genotypic and allelic frequency in a matter of a few seconds. Further usage of SPSS software has been discussed for finding absolute frequencies of various genotypes pertaining to each gene. SPSS also proved to be user friendly menu driven software to obtain descriptive statistics and ANOVA with post - hoc tests for dependent variables pertaining to various genotypes. Final interpretation of ANOVA helped us to determine genes causing significant variation in dependent variables. Thus, the conjunction of HW_TEST and SPSS softwares can successfully be used in statistical analysis of genetic association studies among unrelated individuals.

ABOUT THE AUTHOR



Dr. Kuldeep Kumar Tyagi had completed his B.V.Sc & A.H. in the year 2006 from Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab India. He got admission in a master program in the subject of Animal Genetics and Breeding at Indian Veterinary Research Institute, Bareilly, Uttar Pradesh, India after securing 6th rank in All India ICAR-JRF examination. He had completed his Masters in the year 2008 and carried out research on competent fibroblast cells used in somatic cell nuclear transfer. He qualified CSIR Net in his first attempt during the final semester of the masters program itself. He got selected as Assistant Professor in the year 2009 at College of Veterinary Science & A.H. at Navsari Agricultural University, Navsari, Gujarat, India. He enriched his practical knowledge and expertise in the subject of Animal Breeding while discharging his duties as Scheme Incharge at Livestock Research Station of the same university for 9 years. During the same tenure he also accumulated practical expertise on various aspects of field level breeding programs while heading “All India Coordinated Research Project on Goat Improvement - Surti Field Unit” as Principal Investigator. He completed his Ph.D. in the year 2016 from the same university as an inservice candidate. He had worked on gene expression studies on mammary epithelial cells of buffaloes during his Ph.D. degree program. He had been selected as Associate Professor in the department of Animal Genetics & Breeding, College of Veterinary and Animal Science, Sardar Vallabhbai, Patel University of Agriculture & Technology, Meerut, Uttar Pradesh, India in the year 2018. Since then he has been heading the same department as Officer-Incharge. He had handled 5 externally funded and 27 institutionally funded research projects. He had co-guided two masters students. He has in his credit 62 research papers, 14 research recommendations, 7 lecture notes and 4 success stories. He is a member of 4 professional societies and attended 21 conferences/ symposiums/ workshops. He has remained on a panel of experts for framing question papers for National level, State level examination bodies and various Universities. He is hosting a google site for online teaching <https://sites.google.com/view/learnagb> and can be reached at drtyagivet@gmail.com for initiating a conversation.

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